

**All over the world the lack of screening capacity is a major issue in the SARS-Cov2 pandemy containment. Hereby we share protocols and procedures to help academic institutions and any molecular biology laboratory to provide support to population health care system.**

**In order to guide every laboratory with expertise in molecular biology to develop SARS-CoV2 screening plateform, this starting pack will fasten the changes from basic requirement needed to run RTqPCR test, to build a robust structure.**

### **Contact**

**Pr Nicolas Gillet : [gillet.nicolas@unamur.be](mailto:gillet.nicolas@unamur.be)**

**Pr Patsy Renard : [patsy.renard@unamur.be](mailto:patsy.renard@unamur.be)**

### **Disclaimer**

This document, supported by the University of Namur (hereafter “we,” “us” or “our”), is being distributed in good faith and aims at providing general knowledge for SARS-CoV-2 detection for diagnosis purposes from clinical specimens. This document provides more insights on the protocol as reviewed and accepted by the Belgian AFMPS (Federal Agency for Medicine and Health Products).

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## Starting pack

Requirement



The following protocol requires essential Institution infrastructure, researcher skills and material to ensure efficiency and safe screening procedure



### Institution

- ☐ Biological and chemical waste elimination flow



### Researcher skills

- ☐ Molecular biology manipulation
- ☐ Aseptic experimentation in Biosafety Level 2+ (BL2+)
- ☐ Reverse transcription and quantitative PCR analysis



### Material

- ☐ Class II Biological Safety Cabinet
- ☐ Chemical hood
- ☐ High speed 1,5ml tube refrigerated centrifuges
- ☐ Quantitative PCR machines
- ☐ Micropipettes p1000/p100/p10
- ☐ Centrifuges for 15 mL falcons tubes and qPCR plate
- ☐ -80°C freezer and 4°C fridge



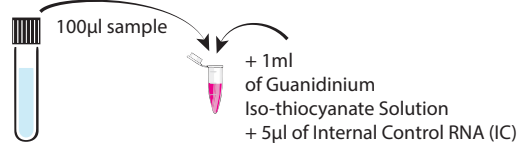
# Starting pack

## Protocol

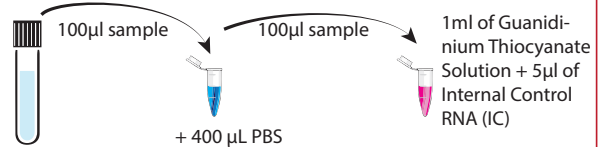


### VIRUS INACTIVATION

#### 1. Sample processing

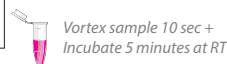


#### REDO with PBS sample 5x dilution

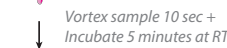


### RNA EXTRACTION

#### 1. Homogenisation



#### 2. Addition of Chloroform

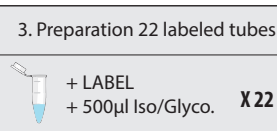
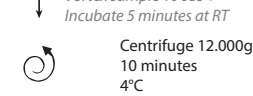


During centrifugation time

#### 4. Transfer RNA aqueous phase



#### 5. Add Isoprop + Glycoblue



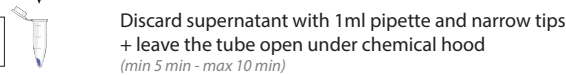
If there is no aqueous phase up



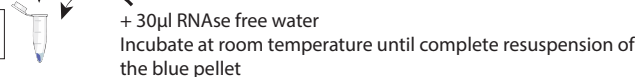
#### 6. Wash the RNA blue pellet



#### 7. Dry the blue pellet



#### 8. RNA resuspension



### one step Reverse Transcript - Quantitative PCR (RTqPCR)

#### 1. Primers and probe mix for SARS and IC

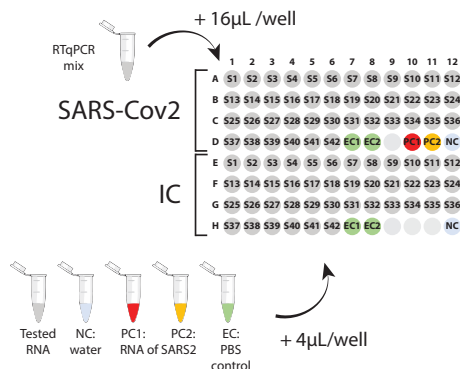
Primers and Probe Mix for SARS  
+ 10 µl of E\_Sarbeco\_Probe [100µM]  
+ 20 µl of E\_Sarbeco\_Fw [100µM]  
+ 20 µl of E\_Sarbeco\_Rev [100µM]  
+ 950 µl of RNAse free water

Primers and Probe Mix for IC (SBV)  
+ 10 µl of IC\_Probe [100µM]  
+ 20 µl of IC\_Fw [100µM]  
+ 20 µl of IC\_Rev [100µM]  
+ 950 µl of RNAse free water

#### 2. RTqCPR mix for SARS and IC

| Volume in µ               | Per reaction | 1 plate | 2 plates | 3 plates | 4 plates | 5 plates |
|---------------------------|--------------|---------|----------|----------|----------|----------|
| SX Master Mix             | 4            | 200     | 400      | 600      | 800      | 1000     |
| EuroscriptII (RT+RNAse I) | 0.2          | 10      | 20       | 30       | 40       | 50       |
| Primers and Probes Mix    | 4            | 200     | 400      | 600      | 800      | 1000     |
| RT Additive               | 0.2          | 10      | 20       | 30       | 40       | 50       |
| RNAse free water          | 7.6          | 380     | 760      | 1140     | 1520     | 1900     |

#### 3. Plate preparation



#### 4. Run

Temperature PCR cycle  
- 48 °C 10 min  
- 95 °C 3 min  
- 45 cycles: 95°C 15 sec 58°C 30 sec

Eurogentec Takyon One-Step RTqPCR



# Starting pack

## Validation and Quality procedure

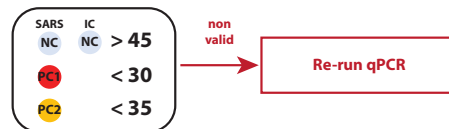


### qPCR result validation

#### Plate

|           |   |     |     |     |     |     |     |     |     |     |     |     |     |
|-----------|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
|           |   | 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  | 12  |
| SARS-Cov2 | A | S1  | S2  | S3  | S4  | S5  | S6  | S7  | S8  | S9  | S10 | S11 | S12 |
|           | B | S13 | S14 | S15 | S16 | S17 | S18 | S19 | S20 | S21 | S22 | S23 | S24 |
|           | C | S25 | S26 | S27 | S28 | S29 | S30 | S31 | S32 | S33 | S34 | S35 | S36 |
|           | D | S37 | S38 | S39 | S40 | S41 | S42 | EC1 | EC2 | PC1 | PC2 | NC  |     |
| IC        | E | S1  | S2  | S3  | S4  | S5  | S6  | S7  | S8  | S9  | S10 | S11 | S12 |
|           | F | S13 | S14 | S15 | S16 | S17 | S18 | S19 | S20 | S21 | S22 | S23 | S24 |
|           | G | S25 | S26 | S27 | S28 | S29 | S30 | S31 | S32 | S33 | S34 | S35 | S36 |
|           | H | S37 | S38 | S39 | S40 | S41 | S42 | EC1 | EC2 |     |     |     | NC  |

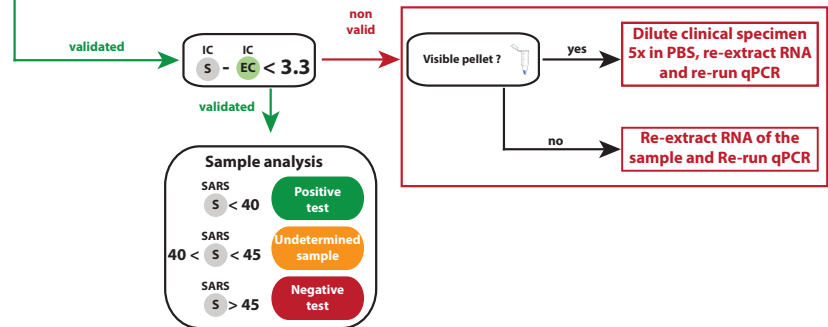
#### Plate validation



#### Extraction batch validation

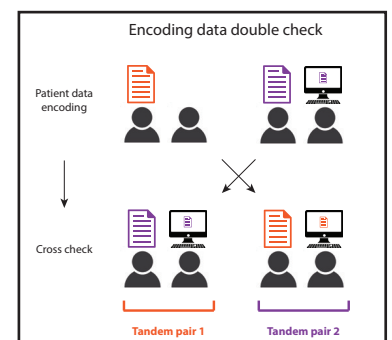
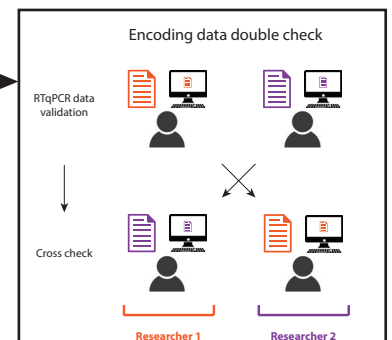
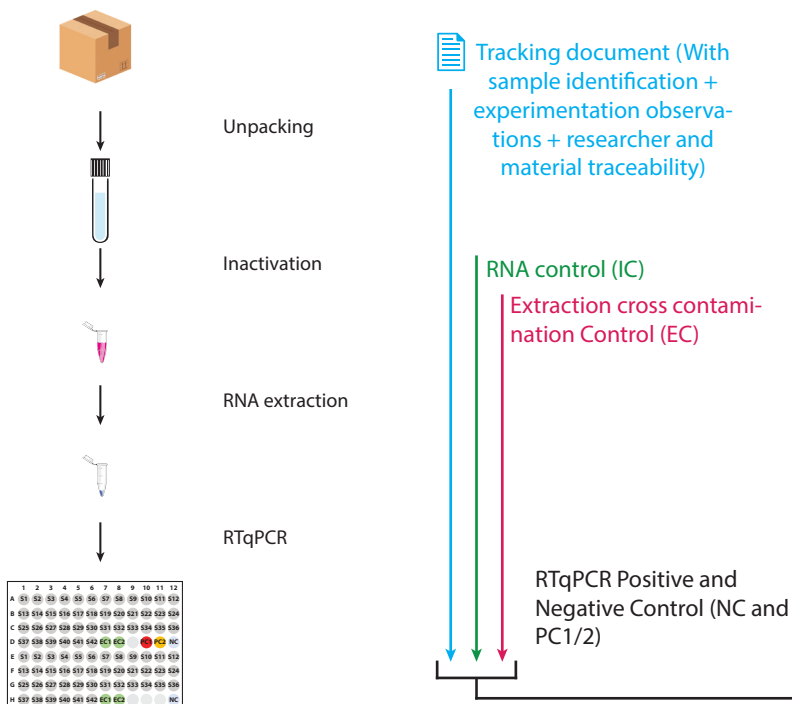


#### Sample validation



### Quality check

#### Process steps

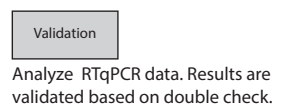
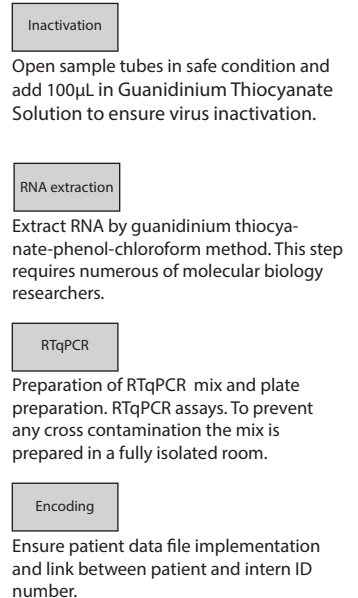




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graph TD; Reception[Reception] --> Tracking[Ensure tracking of intrance samples by intern ID assignation and edit tracability document];
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Reception

Ensure tracking of intrance samples by intern ID assignation and edit tracability document





# Starting pack

Safety advices and sample preservation



## Process steps



Unpacking



Inactivation



RNA extraction



RTqPCR

|   | 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  | 12  |
|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| A | S1  | S2  | S3  | S4  | S5  | S6  | S7  | S8  | S9  | S10 | S11 | S12 |
| B | S13 | S14 | S15 | S16 | S17 | S18 | S19 | S20 | S21 | S22 | S23 | S24 |
| C | S25 | S26 | S27 | S28 | S29 | S30 | S31 | S32 | S33 | S34 | S35 | S36 |
| D | S37 | S38 | S39 | S40 | S41 | S42 | EC1 | EC2 | PC  | NC  |     |     |
| E | S1  | S2  | S3  | S4  | S5  | S6  | S7  | S8  | S9  | S10 | S11 | S12 |
| F | S13 | S14 | S15 | S16 | S17 | S18 | S19 | S20 | S21 | S22 | S23 | S24 |
| G | S25 | S26 | S27 | S28 | S29 | S30 | S31 | S32 | S33 | S34 | S35 | S36 |
| H | S37 | S38 | S39 | S40 | S41 | S42 | EC1 | EC2 |     |     |     | NC  |

## Protective equipment



CLASS II Biological Safety Cabinet



CLASS II Biological Safety Cabinet



Chemical Hood



## Sample preservation

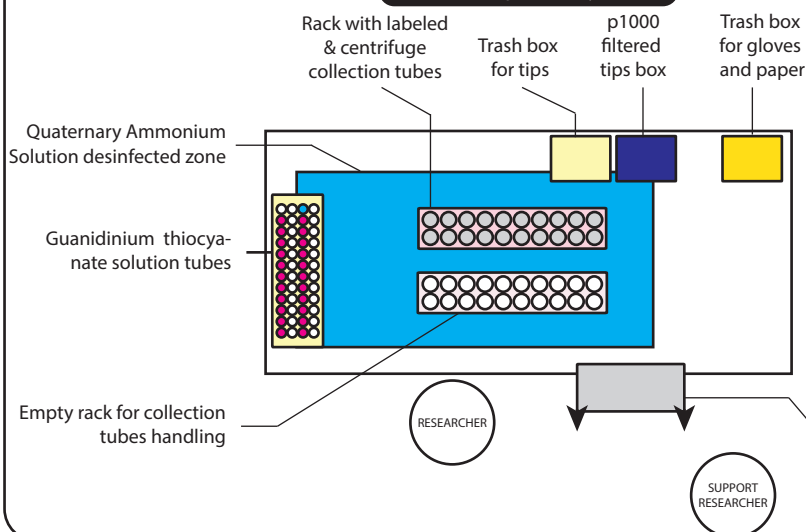
Keep patient sample as much as possible at 4°C

Inactivated samples can be kept at -80°C freezer

Perform RNA extraction in RNase free conditions

Store extracted RNA at 4°C before RTqPCR and next at -80°C for long time conservation

### CLASS II Biological Safety Cabinet



### CLASS II BSF working dress

